- V. V. Vereskovskii and I. I. Chekalinskaya, in: Proceedings of the Third Congress of 2. Pharmacists of the Belorussian SSR [in Russian], Minsk (1977), p. 168.
- 3.
- J. B. Harborne, Biochem. J., <u>70</u>, 22 (1958).
  G. Gorkmann, Phytochemistry, <u>168</u> (1977). 168 (1977). 4.
- K. Hayashi, in: The Chemistry of Flavonoid Compounds (ed. by T. A. Geissman), Pergamon 5. Press, Oxford (1962), p. 248.
- T. K. Chumbalov, G. M. Nurgalieva, and I. D. Beisenova, in: Chemistry and Chemical 6. Technology [in Russian], No. 19, Alma-Ata (1976), p. 146.
- G. M. Nurgalieva and T. K. Chumbalov, in: Phenolic Compounds and Their Biological Func-7. tions [in Russian], Moscow (1968), p. 93.
- Y. Abe and K. Hayashi, Bot. Mag. Tokyo, <u>69</u>, 577 (1956). 8.

INFLUENCE OF TRACE ELEMENTS ON THE ACCUMULATION OF ANTHOCYANINS IN THE FRUIT OF Aronia melanocarpe

E. G. Martynov

UDC 547.56

It has been established that in addition to vitamins, catechins, and flavonoids, the active substances of the fruit of Aronia melanocarpa Elliot. (black chokeberry) are anthocyanins - substances possessing a high biological activity [1].

The aim of our work was to study the accumulation of anthocyanins in the ripe fruit of the black chokeberry under the influence of trace elements and to investigate their qualitative composition.

Foliar feeding was carried out with 0.03% solutions of: H<sub>3</sub>BO<sub>3</sub>; ZnSO<sub>4</sub>·7H<sub>2</sub>O; CoSO<sub>4</sub>·7H<sub>2</sub>O;  $CuSO_4 \cdot 5H_2O$ ;  $(NH_4)_2MoO_4$ ;  $FeSO_4 \cdot 7H_2O$ ; and  $MnSO_4 \cdot nH_2O$  by a method described previously [2] in the "Kiritsy" sovkhoz [communal farm] in the Spassk region of the Ryazan oblast and at the young naturalists's mountain station.

The anthocyanin substances were isolated from the crude ripe fruit with 1% HCl in methanol (1:99) [3]. The extract was evaporated in vacuum and was studied by partition paper chromatography (Leningrad type "M" ["slow"] paper) in the n-butanol-acetic acid-water (4:1:5) system. In all variants of the experiment three anthocyanin substances were detected, having  $\lambda_{\text{max}}^{\text{CH}_{3}\text{OH}}$  + 1% HCl 531, 535, and  $R_{f}$  0.35, 0.44, and 0.67 and with the respective UV spectra: 537 nm [4]. After the hydrolysis of these substances with 2 N HCl for 30 min [5] and rechromatography of the hydrolysis products isolated by isoamyl alcohol, the three anthocyanin substances formed one and the same aglycone, which was identified as cyanidin, having  $R_f$  0.62.

 $\lambda_{\text{max}}^{C_2H_5OH}$  545 nm and  $\lambda_{\text{max}}^{CH_3OH}$  + 1% HCl 542 nm [3, 4]. UV spectra:

The amounts of the anthocyanins in the ripe fruit of the 1973-1976 harvests were determined as cyanidin by the photocolorimetric method [6]. The results of the analysis (in % of the dry matter) were treated by the statistics of variations [7].

It was established that Co, Zn, B, Mn, Mo, Cu, and B + Zn increase the accumulation of anthocyanins by 7, 9, 11, 12, 13, 19, and 20%, respectively.

## LITERATURE CITED

- 1. M. N. Zaprometov, in: Vitamin Resources and Their Utilization [in Russian], No. 4, Moscow (1959), p. 6.
- E. G. Martynov and N. I. Suprunov, in: Question of the Development of New Drugs [in 2. Russian], No. 50, Ryazan (1975), p. 16.
- L. O. Shnaidman, V. S. Afanas'eva, and O. P. Khorina, in: Proceedings of a Conference 3. on Vitamins from Natural Raw Materials [in Russian], Kuibyshev (1964), p. 156.
- 4. A. A. Kolesnik and L. G. Elizarova, in: Phenolic Compounds and their Biological Func-

I. P. Pavlov Ryazan Medical Institute. Translated from Prirodnykh Soedinenii, No. 4, p. 526, July-August, 1978. Original article submitted May 3, 1978.

tions [in Russian], Moscow (1968), p. 233.

- 5. G. B. Samorodova-Bianki, Fiziologiya Rastenii, 11, No. 3, 544 (1964).
- 6. L. O. Shnaidman and V. S. Afanas'eva, in: Ninth Mendeleev Congress on General and Applied Chemistry [in Russian], Moscow, No. 7 (1965), p. 79.
- 7. P. F. Rokitskii, in: Biological Statistics [in Russian], Minsk (1973), p. 320.

PROANTHOCYANIDINS OF THE LEAVES OF Laurocerasus officinalis

G. Ch. Tsiklauri

UDC 547.972

We have previously reported the isolation from the leaves and flowers of *Laurocerasus* officinalis Roem. (Prunus laurocerasus; common laurel cherry) of catechins and anthocyanidins and their identification [1, 2]. Continuing our investigation of the leaves of this plant, we have isolated two more compounds (I, II) by column chromatography on cellulose. Elution was performed with distilled water [3].

The compounds obtained gave the qualitative reactions characteristic for leucoanthocyanidins with the vanillin reagent and with p-toluenesulfonic acid, and on being heated with the leucoanthocyanidin reagent they were converted into the corresponding anthocyanidins [4, 5].

In the butan-1-ol-acetic acid-water (40:12:28) solvent system, compounds (I) and (II) has  $R_f 0.55$  and 0.50, and in 2% acetic acid 0.44 and 0.46, respectively. The UV spectra [ $\lambda_{max}$  (ethanol), 280 nm] and IR spectra [3400-3000 cm<sup>-1</sup> (OH), 1615-1600 cm<sup>-1</sup> (C<sub>6</sub>H<sub>5</sub>), and 1000 cm<sup>-1</sup> (C-C)] of the two compounds were identical.

The anthocyanidins obtained from the compounds (I) and (II) by heating with the leucoanthocyanidin reagent were studied chromatographically and spectrally. They had  $R_f$  0.58 in the acetic acid-hydrochloric acid-water (30:35:10) solvent system and 0.32 in the same system at a ratio of 5:1:5. The absorption maxima in the visible region of the spectrum were also identical:  $\lambda_{max}$  (ethanol) 545 nm, (+ AlCl<sub>3</sub>) 562 nm. These results agree with the chromatographic and spectral characteristics of authentic cyanidin.

After acid hydrolysis [6] of the initial compounds, (+)-catechin for compound (I) and (+)-catechin and (-)-epicatechin for compound (II) were found in the hydrolysates by paper chromatography with markers.

In the study of the products of alkaline cleavage [7] of compounds (I) and (II), phloroglucinol and protocatechuic acid were detected, being identified by comparison with authentisamples on paper chromatography.

On the basis of the results obtained it has been established that compound (I) is a dimer consisting of (+)-catechin and leucocyanidin and (II) is a trimer the monomers of which are (+)-catechin, (-)-epicatechin, and leucoanthocyanidin. The ease of liberating the monomers on the acid hydrolysis of (I) and (II) with dilute acid and literature information show that the monomers are linked through a  $C_4$ - $C_8$  bond [7-9].

## LITERATURE CITED

G. Ch. Tsiklauri and A. G. Shalashvili, Soobshch. Akad. Nauk GSSR, 77, No. 3, 721 (1975 1. G. Ch. Tsiklauri, Soobshch. Akad. Nauk GSSR, 79, No. 1, 177 (1975). 2. W. R. Chan. W. G. Forsyth, and C. H. Hassal, J. Chem. Soc., 3174 (1958). 3. D. G. Roux and S. R. Evelyn, Biochem. J., <u>69</u>, No. 4, 530 (1958). 4. T. Swain and W. E. Hillis, J. Sci. Food Agric., 10, 63 (1959). 5. S. Lewak, J. Lazowska, and K. Dziewanowska, Roczniki Chem., 44, No. 9, 1733 (1970). 6. N. D. Storozhenko and T. K. Chumbalov, Khim. Prirodn. Soedin., 735 (1976). 7. S. Rangeswami and P. Vencateswarlu, Proc. Indian Acad. Sci., A 64, No. 3, 185 (1966). 8. E. Haslam, Phytochemistry, 16, 1625 (1977). 9.

Institute of Plant Biochemistry, Academy of Sciences of the Georgian SSR, Tbilisi. Translated from Khimiya Prirodnykh Soedinenii, No. 4, pp. 526-527, July-August, 1978. Original article submitted May 3, 1978.